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(54) **Albumin-based nucleotides, their replication and use, and plasmids for use therein.**

(57) The DNA sequence coding for human serum albumin has been isolated and inserted as two fragments into two novel plasmids which can be replicated in *E. coli*. These novel fragments can be joined to provide a unitary DNA sequence which then can be cloned into a suitable host, e.g. *E. coli*, for the expression of human serum albumin (which is used extensively in medical practice in treating shock conditions).

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ALBUMIN-BASED NUCLEOTIDES, THEIR REPLICATION
AND USE, AND PLASMIDS FOR USE THEREIN

This invention relates to nucleotides related to human serum albumin (HSA), their replication and use, and plasmids (and host substances) for use therein.

The gene for serum albumin is regulated in

5 development. On the other hand, serum albumin is synthesised in mammals by the adult liver, and its plateau in adulthood. The embryonic liver and yolk sac, on the other hand, produce predominantly α -fetoprotein, but the synthesis decreases drastically after birth. Recently, 10 Law et al determined the complete sequence of mouse α -fetoprotein mRNA, Nature 291 (1981) 201-205. The structure revealed extensive homology to mammalian serum albumin, indicating that the two proteins are encoded in the same gene family. Similar conclusions have been 15 reached from studies on the α -fetoprotein genes of the rat and the mouse; see Jagodzinski et al, Proc. Natl. Acad. Sci. USA, 78 (1981) 3521-3525, and Gorin et al, J. Biol. Chem. 256 (1981) 1954-1959.

The complete nucleotide sequence of human serum 20 mRNA has been determined from recombinant cDNA clones and from a primer-extended cDNA synthesis on the mRNA template. The sequence comprises 2,078 nucleotides, starting upstream of a potential ribosome binding site in the 5'-untranslated region. It contains all the 25 translated codons and extends into the poly(A) at the 3'-terminus. Part of the translated sequence codes for a hydrophobic prepeptide met-lys-trp-val-thr-phe-ile-ser-leu-leu-phe-leu-phe-ser-ser-ala-tyr-ser, followed by a basic propeptide arg-gly-val-phe-arg-arg. These signal 30 peptides are absent from mature serum albumin and, so far, have not been identified in their nascent state in humans. A remaining 1,755 nucleotides of the translated mRNA sequence code for 585 amino acids which are in agreement, with few exceptions, with the published amino 35 acid data for human serum albumin. The mRNA sequence verifies and refines the repeating homology in the triple-domain structure of the serum albumin molecule.

DETAILED DESCRIPTION OF THE INVENTION

Human serum albumin cDNA is cloned into the PstI site of plasmid pBR322 by the oligo(dG)-oligo(dC) tailing technique. Plasmid DNA was isolated from 97 positive colonies which hybridized to the enriched 5 albumin cDNA probe, and the recombinant plasmid pH A36 was found to contain the largest insert of an albumin cDNA sequence. Its restriction endonuclease map is shown in the drawing, together with a restriction map of the primer-extended plasmid clone pH A206. The latter was obtained in a second transformation experiment after initiating 10 the cDNA synthesis from an internal primer. This primer was a 91 base pairs long DNA fragment, MspI(152)-TagI(182/3), isolated from pH A36. The two plasmids, pH A36 and pH A206, share 0.15 kb of homologous DNA. Together, they encode the entire sequence for human serum albumin, starting with the CTT codon for leu -10 of the prepeptide and extending 15 into the 3'-untranslated region of poly(A).

Sequence of the Albumin cDNA. The sequence was determined for the most part on both DNA strands to ensure accuracy. All of the restriction sites used to end-label DNA fragments were sequenced across by 20 labeling a neighboring restriction site. The entire nucleotide sequence of the serum albumin mRNA, as determined from the cloned DNA in pH A36, pH A206, and from the primer-extended cDNA at the 5'-terminus of the message, is shown in the following Table 1. The inferred amino acid sequence is also indicated. The mRNA length is 2,078 nucleotides, of which 38 represent the 5'-untranslated region, 54 identify a 25 prepeptide of 18 amino acids, 18 identify a propeptide of 6 amino acids, 1,755 code for the known 585 amino acids of serum albumin, 189 make up the 3'-untranslated region and 24 are the poly(A) sequence. Nucleotides 5 to 15 (-34 to -24) in the 5'-untranslated region (Table 30 1) are complementary to a 3'-terminal region of eukaryotic 18S RNA [Azad, A.A. and Deacon, N.J. (1980) Nucl. Acids Res. 8, 4365-4376] and thus could represent a ribosome binding site:

(5')...T T^C T C T T C T G T.....albumin mRNA
35 (3')...G A G G A A G G C G U C C m₂⁶A m₂⁶A.....18S RNA

The translated portion of the mRNA sequence codes for the signal peptide and the main body of the albumin polypeptide chain. The

signal peptide is composed of a hydrophobic prepeptide of 18 amino acids and a basic propeptide of 6 amino acids (Table 1). Since pre-peptides are removed from nascent secretory proteins (like albumin) in the endoplasmic reticulum, they are seen only in vitro in heterologous 5 translation systems. As yet, they have not been found within cells [Judah, J.D. and Quinn, P.S. (1977) FEBS 11th Mtg., Copenhagen 50, 21-29; and Strauss, A.W., Donohue, A.M., Bennett, C.D., Rodkey, J.A. and Alberts, A.W. (1977) Proc. Natl. Acad. Sci. USA 74, 1358-1362]. This is the first report of the presence and the sequence of a pre- 10 peptide for human serum albumin. As it is with other secretory proteins, the conversion of proalbumin to albumin takes place in the Golgi vesicles, and the enzyme responsible for this cleavage is probably cathepsin B [Judah, J.D. and Quinn, P.S. (1978) Nature 271, 384-385]. This is also a first report on the sequence of the pro- 15 peptide for normal human serum albumin.

At the 3'-end of the message, the putative polyadenylation signal sequence, AATAAA, is located 164 nucleotides downstream from the amino acid termination codon TAA and 16 nucleotides upstream from the beginning of the poly(A) sequence. Another characteristic sequence 20 located near the polyadenylation site has been identified by Benoist, et al. [Benoist, C., O'Hare, K., Breathnach, R. and Chambon, P. (1980) Nucl. Acids Res. 8, 127-142]; the consensus sequence from several mRNAs was concluded as TTTTCACTGC. A similar sequence, TTTTCTCTGT, is located 19 nucleotides upstream from the AATAAA hexanucleotide in the 25 human albumin mRNA (Table 1).

TABLE 1

5 10 15 20 25 30 35

-10 -5

-18 p r o -10

Met lys trp val thr phe ile ser leu leu phe leu ala ser
GCTTTCCTCTGTCAACCCACACCCCTTTGCCACA ATG AAC TGG GAA CTC ATT TCC CTT CTT TTT CTC TTT AGC (30)

-1 -6 p r o -1 1

ser ala tyr ser arg gly val phe arg asp ala his lys ser glu val ala his arg phe lys asp leu ala glu glu asn phe lys
TCC CCT TAT TCC ACC CGT GTC TTT CGT CCA GAT CCA AAC ACT GAG CCT GCT CAT CGT CAA GAT CAT GAA GAT CAA GAT CAA ACT CAA TTT GCA (170)

21 30 34 40 50

ala leu val lle ala phe ala gln tyr leu gln gln cys pro phe glu asp his val lys leu val asn glu val thr glu phe ala
GCC TTG CTG TTG ATT GCC TTT CCT CAG TAT CTT CAG CAG TGT CCA TTT GAA GAT CAT GAA GAT CAA TAA TTA GTC AAC AAT GAA GAT CAA ACT CAA TTT GCA (260)

51 53 60 62 70 75 80

lys thr cys val ala asp glu ser ala glu asn cys asp lys ser leu his thr leu phe gly asp lys leu cys thr val ala thr leu
AAA ACA TCT CCT CCT GAT GAG TCA CCT GAA ATT TGT GAC AAA TCA CTT CAT ACC CCT TTT CGA GAC AAA TTA TGC ACA CCT CCT CAA ACT CCT (350)

81 90 91 100 101 110

arg glu thr tyr gly glu met ala asp cys cys ala lys gln glu pro gly arg asn glu oys asn ala glu qin his lys asp asp asn pro
CGT GAA ACC TAT CCT GAA ATG CCT GCA TGT GCA AAC CAA CAA CCT GCG ACA AAC TAA TGC TTC TTG CAA CAC AAA GAT GAC AAC CCA (440)

111 120 124 130 140

asn leu pro arg leu val arg pro glu val asp val met cys thr ala phe his asp asn glu glu thr phe leu lys tyr leu try
AAC CTC CCC CGA TTG CTG AGA CCA GAG CCT GAT CTC ATG ACT CCT TTT CAT GAC AAC TAA GAT GAA GAG TAT AAA AGG TAT AAA TAC TTA TAT (330)

141 150 160 168 169 170

glu lle ala arg arg his pro tyr phe tyr ala pro glu leu leu phe ala lys arg tyr lys ala phe thr glu cys cys qin
GAA ATT GCC AGA AGA CAT CCT TAC TTT TAT GCC CCC GAA CTC CCT CCT TTT GCT GAA ATT CCT GCA CTC CCT GAA GAG CCT TCC TCT CCC AAA CAG AGA CTC AGC TGT GCA (620)

171 177 180 190 200

ala ala asp lys ala ala cys leu leu pro lys leu arg asp glu leu arg asp glu ala ser ser ala lys qin arg leu lys cys
GCT CCT GAT AAA CCT CCT CCC TGC TGC CCA AAC CTC GAT GAA CCT CCT GAT GAA GAG CCT TCC TCT CCC AAA CAG AGA CTC AGC TGT GCA (710)

201 210 220 230

ala ser leu gln lys phe gly glu arg ala phe lys ala trp ala val ala arg leu ser gln arg phe pro lys ala glu phe ala glu
GCC AGT CTC CAA AAA TTT GCA GAA AGA GCT TTT CCC AAA GCT CCT CTC AGC CAG AGC TTT GCA GAA (300)

231 val ser lys leu val thr asp leu thr lys val his thr glu cys oys his gly asp leu glu cys ala asp arg ala asp leu
 GTC TCC AAC TTA GTC ACA GAT CCT ACC AAA GTC CAC ACC GAA TCC TGC CTT GCA GAT CTC CCT GAA TGT GCT GAT GAC AGC CCC GAC CTT (890)
 261 ala lys tyr lle cys glu asn gln asp ser lle ser ser lys leu lys glu oys cys glu lys pro leu leu glu lys ser his cys lle
 GGC AAC TAT ATC TGT GAA AAT CAA GAT TCG ATC TCC ACT AAA CTG AAC GAA TCC TGT GAA AAA CCT CTC TTC GAA AAA TGT GAT ATT (980)
 291 ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala ala asp phe val glu ser lys asp val cys lys ASN tyR tyR ala
 GGC GAA GTC GAA AAT GAT GAG ATG CTC CCT GCT GAC TTG CCT TCA TTA GCT GAT TTT GTC GAA ACT AAC CAT GTT TGT AAA AAC TAT GCT GTC (1070)
 321 glu ala lys asp val phe leu gly met phe leu tyr ala ala arg his pro esp tyR ser val val phe asp glu phe lys pro leu ala
 GAG GCA AAG GAT GTC TTC TTG GGC ATG TTT TGT TAT GAA TAT GCA AGA ACC CAT CCT GAT TAC TCT GTC CTC CTC (1160)
 351 lys thr tyr glu thr thr leu glu lys cys cys ala ala ala asp pro his glu cys tyR ala lys val phe asp glu phe lys pro leu
 AAC ACA TAT GAA ACC ACT CTA GAG AAC TGC CCT GCT GTC TGT GCA GAT CCT GCA GAT CTC TAT GCA TGT GAA TTT AAA CCT CCT (1250)
 381 val glu glu pro gln asn leu lle lys gln asn cys glu leu phe glu qln leu qly glu tyR lys phe gln asn ala leu leu val arg
 GTC GAA GAG CCT CAG AAC TTA ATC AAA AAC CAA GTC TCA ACT CCT GCA ACT CTT GTA GAC GTC TCA AGA AAC CTA GGA AAA TGT TGT AAA CAT (1340)
 411 tyr thr lys lys val pro gln val ser arg asn leu gln lys val gln lys ser lys cys cys lys his
 TAC ACC AAC AAA GTA CCC CAA GTC TCA ACT CCT GCA ACT CTT GTA GAC GTC GTC AAC CAC TCA TCC TGT GAT GAA ACA TAC GTC GAA AAA TGT TGT AAA CAT (1430)
 441 pro glu ala lys arg met pro oys ala glu asp tyR leu ser val val asn gln leu cys val lys thr pro val ser
 CCT GAA GCA AAA ACA ATG CCC TGT GCA GAA GAC TAT CTA TCC GTC CTC AAC CAC TCA TCC TGT GAT GAA ACA TAC GTC GAA CCC GCA ACT CCT GTC (1520)
 471 asp arg val thr lys oys cys thr glu ser leu val asn arg pro cys phe ser ala leu glu val asp glu thr tyR val pro lys
 GAC AGA GTC ACC AAA TGC TGC ACA GAA TCC TTG GTC AAC ACC CCA CCA TGC CCT CTC GAA GTC GAT GAA ACA TAC GTC GAA CCC AAA (1610)
 501 glu phe asn ala glu thr phe thr phe his ala asp lle cys thr leu ser glu lys glu arg gln thr ala leu val
 GAG TTT ATT CCT GAA ACA TTC ACC FTC CAT CCA GAT ATT TGC ACA CTT TCT GAC AAC GAG CAA ATC AAC CAA ACT GCA CTT CCT (1700)

Following are examples which illustrate procedures, including the best mode, for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

5 Example 1 Isolation of Messenger RNA

Human liver mRNA was obtained following the procedure of Chirgwin, et al [Chirgwin, J.M., Przybyla, A.E., MacDonald, R.J. and Rutter, W.J. (1979) Biochemistry 18, 5294-5299]. Immunoprecipitation of albumin containing polysomes was performed according to Taylor and 10 Tse [Taylor, J.M. and Tse, T.P.H. (1976) J. Biol. Chem. 251, 7461-7467]. In vitro translation of mRNA was carried out in a reticulocyte cell-free system, following the instruction of the manufacturer (New England Nuclear). The translation products were separated electrophoretically according to Laemmli [Laemmli, J.K. (1970) Nature 227, 15 680-685.

15 Example 2 Cloning Procedures

Double stranded cDNA was synthesized as described previously [Law, S., Tamaoki, T., Kreuzaler, F. and Dugaiczyk, A. (1980) Gene 10, 53-61]. It was annealed to PstI-linearized pBR322 DNA [Bolivar, F., 20 Rodriguez, R.L., Greene, P.J., Betlach, M.C., Heyneker, H.L., Boyer, H.W., Crossa, J.H. and Falkow, S. (1977) Gene 2, 95-113] that had been tailed with 15 dG residues/3'-terminus [Dugaiczyk, A., Robberson, D.L. and Ullrich, A. (1980) Biochemistry 19, 5869-5873]. The annealed DNA was used to transform E. coli strain RR1, as detailed previously [Law, 25 S., et al., Ibid.]. The albumin clones were selected using the colony hybridization method of Grunstein and Hogness [Grunstein, M. and Hogness, D.S. (1975) Proc. Natl. Acad. Sci. USA 72, 3961-3965], with [³²P]-labeled cDNA synthesized with the immunoprecipitated polysomal mRNA as template.

30 As shown in Example 5, plasmids pH A36 and pH A206 were deposited in E. coli HB101 hosts. The plasmids were obtained from E. coli RR1 hosts, described in this example, and transformed into E. coli HR101 by standard procedures well known to those of ordinary skill in this art. The E. coli RR1 hosts were lysed and then centrifuged to 35 separate the chromosomal DNA, cell DNA and plasmid DNA. The plasmid DNA, remaining in the supernatant, is precipitated with ethanol and the precipitate is resuspended in buffer, e.g., TCM (10mM Tris-HCl, pH 8.0, 10 mM CaCl₂, 10 mM MgCl₂). The cells for transformation are

prepared as follows: 120 ml of L-broth (1% tryptone, 0.5% yeast extract, 0.5% NaCl) are inoculated with an 18 hour culture of HR101 NRRL B-11371 and grown to an optical density of 0.6 at 600 nm. Cells are washed in cold 100 mM NaCl and resuspended for 15 minutes in 20 ml 5 chilled 50 mM CaCl₂. Bacteria are then concentrated to one-tenth of this volume in CaCl₂ and mixed 2:1 (v:v) with annealed plasmid DNA, prepared as described above. After chilling the cell-DNA mixture for 15 minutes, it is heat shocked at 42°C for 2 minutes, then allowed to equilibrate at room temperature for ten minutes before addition of 10 L-broth 10 times the volume of the cell-DNA suspension. Transformed cells are incubated in broth at 37°C for one hour before inoculating selective media (L-agar plus 10 µg/ml tetracycline) with 200 µl/plate. Plates are incubated at 37°C for 48 hours to allow the growth of transformants.

15 Example 3 Mapping of Restriction Endonuclease Sites

Restriction endonucleases were obtained from Bethesda Research Laboratories and New England Biolabs and were used according to the manufacturers' instructions. The digested DNA fragments were analyzed electrophoretically on agarose [Helling, R.B., Goodman, H.M. and 20 Boyer, H.W. (1974) J. Virol. 14, 1235-1244] or acrylamide [Dingman, C., Fisher, M.P. and Kakefuda, T. (1972) Biochemistry 11, 1242-1250] gels.

Example 4 DNA Sequencing

DNA fragments were dephosphorylated with bacterial alkaline 25 phosphatase (Worthington) and labeled at the 5'-ends with poly-nucleotide kinase (Boehringer-Mannheim) and γ [³²P]ATP. Following digestion with a second restriction endonuclease and electrophoretic separation of the fragments, DNA sequence determination was done according to the procedure of Maxam and Gilbert [Maxam, A. and 30 Gilbert, W. (1980) Methods Enzym. 65, 499-560] and the degradation products were separated electrophoretically on 0.4 mm acrylamide gels as described by Sanger and Coulson [Sanger, F. and Coulson, R. (1978) FEBS Letters 87, 107-110].

Example 5 Recombinant Plasmids pH A36 and pH A206

35 As disclosed in Example 2, albumin clones were selected by hybridizing to the enriched albumin cDNA probe. Plasmid pH A36 contained the largest insert of an albumin cDNA sequence. Both plasmids pH A36 and pH A206 have been deposited in a viable E. coli host in the

permanent collection of the Northern Regional Research Laboratory (NRRL), U.S. Department of Agriculture, Peoria, Illinois, U.S.A. Their accession numbers in this repository are as follows:

HB101(pHA36) - NRRL B-12551

5 HB101(pHA206) - NRRL B-12550

E. coli HB101 is a known and widely available host microbe. Its NRRL accession number is NRRL B-11371.

NRRL B-12550 and NRRL B-12551 are available to the public, upon the grant of a patent. It should be understood that the availability 10 of these deposits does not constitute a license to practice the subject invention in derogation of patent rights granted with the subject instrument by governmental action.

E. coli RR1 and E. coli HB101 are known and widely available host microbes. Their NRRL accession numbers are NRRL B-12186 and NRRL 15 B-11371, respectively.

pBR322 is a well known and widely available plasmid. It can be obtained from the following host deposit by standard procedures:

NRRL B-12014 - E. coli RR1 (pBR322).

YEp6 is a well known and widely available yeast episomal plasmid. 20 It can be obtained from the following host deposit by standard procedures:

E. coli HB101 (YEp6) - NRRL B-12093.

Example 6 Assembly of the Serum Albumin Gene

Assembling the pieces together is a straightforward task of restriction enzymology. There is only one MspI site in the overlapping 25 DNA sequence of the two cDNA clones. Two enzymatic steps of (i) MspI digestion of the two DNAs, followed by (ii) the use of ligase, an enzyme that seals DNA fragments, will give the desired product. Although two other undesired DNA species will also be obtained in the 30 course of this recombination reaction, both of them will differ substantially in size. Thus, separation and isolation of the desired DNA species will be achieved.

The assembled DNA clone can be used to transform two types of cells:

35 (a) Escherichia coli

(b) Saccharomyces cerevisiae

(a) The vector of choice is plasmid pBR322, the same that has

been successfully used for cloning of the two fragmented pieces of the serum albumin cDNA.

(b) In order to transform yeast with the serum albumin structural gene sequence, the DNA must be inserted into one of the 5 existing yeast plasmid vectors. This can be accomplished by taking advantage of the fact that several restriction endonuclease recognition sequences are absent from the cloned serum albumin DNA. Synthetic EcoR1 DNA linkers can be ligated to the DNA fragment containing the serum albumin sequence followed by insertion (ligation) into one 10 of the yeast plasmid vectors, e.g., YEp6, at the EcoR1 cloning site. The fused chimeric plasmid can be used to transform yeast according to an established procedure [Hinnen, A., Hicks, J.B. and Fink, G.R. (1978) Proc. Natl. Acad. Sci. USA, 75, 1929]. YEp6 can be obtained from the NRRL repository, as disclosed *supra*.

15 Example 7 Expression of the Serum Albumin Gene

The main body of the structural gene will be transcribed by the E. coli or yeast enzymes. If little or no albumin is produced with the selected host, then an Escherichia coli promoter DNA sequence carrying an initiation codon, i.e., ATG, can be ligated at the beginning 20 of the serum albumin structural gene. Such elements are known and available, e.g., lac promoter used for the expression of human interferon gene in E. coli [Proc. Natl. Acad. Sci. 77, 5230 (1980)]; source of promoter DNA [Proc. Natl. Acad. Sci. 76, 760 (1979)]. Also, see Nature, Vol. 281, October 18, 1979. It has already been 25 documented that such Escherichia coli promoter sequences function well in the expression of foreign genes in Escherichia coli [Mercereau-Puijalon, O., Royal, A., Cami, B., Garapin, A., Krust, A., Gannon, I. and Kourilsky, P. (1978) Nature 275, 505; and Goeddel, D.V., Kleid, D.G., Bolivar, F., Heyneker, H.L., Yansura, D.G., Grea, R., Hirose, 30 T., Kraszewski, A., Itakura, K., and Riggs, A. (1979) Natl. Acad. Sci. USA 76, 106]. For expression in yeast, see Rose, M., Casadaban, M.J. and Botstein, D. (1981) Proc. Natl. Acad. Sci. USA 78, 2460 and 4466.

Example 8 Screening of Clones Producing Albumin

35 Immunological methods can be used to detect small amounts of albumin made in a bacterium. Flat disks of flexible polyvinyl are coated with the IgG fraction from an immune serum and the disks are pressed onto an agar plate so that antigen released from an in situ lysed microbial colony can bind to the fixed antibody. The plastic

disk is then incubated with the same total IgG fraction labeled with radioactive iodine so that other determinants on the bound antigen can in turn bind the iodinated antibody. Radioactive areas on the disk expose X-ray film during autoradiography and thus identify colonies producing the protein which is being screened for. Detailed protocols of this procedure have been published [Broome, S. and Gilbert, W. 5 (1978) Proc. Natl. Acad. Sci. USA, 75, 2746]. The purification of human serum albumin can be accomplished by using procedures well known in the art. For example, procedures disclosed in a chapter by T. 10 Peters: Purification and Properties of Serum Albumin, in: The Plasma Proteins, Putnam, Ed. Academic Press, New York, 1975, can be used.

The work described herein was all done in conformity with physical and biological containment requirements specified in the NIH Guidelines.

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CLAIMS

1. Plasmid pH A36, having a restriction endonuclease pattern as shown in the drawing.

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2. Plasmid pH A206, having a restriction endonuclease pattern as shown in the drawing.

3. E. coli HB101 (pHA36) having the deposit accession number
10 NRRL B-12551.

4. E. coli HB101 (pHA206) having the deposit accession number
NRRL B-12550.

15 5. A microorganism modified to contain a nucleotide sequence coding for the amino acid sequence of human serum albumin; said nucleotide sequence is as follows:

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0079739

4083

-13-

0079739

4083

-15-

6. Nucleotide sequence of the cDNA of human serum albumin, said nucleotide sequence is as follows:

35 30 25 20 15 10 5

231 240 245 246 250 253 260
val ser lys leu val thr asp leu thr lys val his thr glu cys his gly asp leu glu cys ala asp asp arg ala asp leu
GTT TCC AAC GAA CAT CTT ACC AAA GTC CAC ACC GAA TGC TCG CAT GAA CTT GCA GAT GTC CTC TGA TGT CCT GAT GAC AGC CCC GAC CTT (690)

261 265 270 278 279 280 289 290
ala lys tyr ile cys glu asn gln asp ser ile ser ser lys leu lys glu cys glu lys pro leu glu cys ser his cys ile
CCC GAA GTC GAA AAT CAT GAC ATG CCT CCT GCT GAT TTA CCT TCA TTA GCT GAA AGT AAC GAT GTT TGC AAA AAC TAT GCT (980)

291 300 310 316 320 328 340 350
ala glu val glu asn asp glu met pro ala asp leu pro ser leu ala asp phe val glu ser lys asp val cys lys asn tyro ala
CCC GCA GTC GAA AAT CAT GAC ATG CCT CCT GCT GAT TTT GCT GAA AGT AAC GAT GTT TGC AAA AAC TAT GCT (1070)

321 330 340 348 350
glu ala lys asp val phe leu gly met phe leu tyr ala arg arg his pro asp tyro ser val val leu leu arg leu ala
GAG CCA AAC GAT GTC TTC TTC GGC ATG TTT TGC TAT GCA TAT GCA AGA AGC CAT CCT GAT TAC TCT GTC GTC CTC CTC GCA ACT CCC (1160)

351 360 361 369 370 380 390 392 400 410
lys thr tyr glu thr thr leu glu lys cys ala ala asp pro his glu cys tyr ala lys val phe asp glu phe lys pro leu
AAG ACA TAT GAA ACC ACT CTA GAG AAC TGC TGT GCC CCT GCA GAT CCT CAT CAA TGC TAT GCA AAA GTC TTT AAA CCT CCT CGT (1250)

381 390 392 398 400 410
val glu glu pro gln asn leu lys gln asn cys glu leu phe glu aln leu gln glu tyro lys phe gln asn ala leu val arg
GTC CAA GAC CCT CAG AAC TTA ATC AAA CAA ATT TGT GAC CTT TTT GAG CAG CCT CTC GAG TAC AAA TTC CAC AAT GGC CTC TTA GTC CGT (1340)

411 420 430 437 438 440
tyr thr lys lys val pro gln val ser thr pro thr leu val glu val ser arg asn leu gln lys val gln ser lys cys oys his
TAC ACC AAG AAA GTC CAA CCC CAA GTC TCA ACT CTT GTA AAC CAG GTC TCA AGA AAC CTA GCA AAA GTC CGC AGC ACC AAA TGT TGT AAA CAT (1430)

441 448 450 460 461 470
pro glu ala lys arg met pro oys ala glu asp tyro leu ser val val gln leu cys val his glu lys thr pro val ser
CCT GAA GCA AAA AGA ATG CCC TGT GCA GAA GAC TAT CTA TCC AAC AGC GTC CTC AAC CAG TTA TGT GTC CAT GAA ACA TAC GTC CCC GAA ACT (1520)

471 476 477 480 490 500
asp arg val thr lys cys cys thr glu ser leu val arg arg pro cys phe ser ala leu glu val asp glu thr tyro val pro lys
GAC ACA GTC ACC AAA TGT GCA GAA ACA TCC AAC AGC GTC CTC AAC CAG TGA TCT GCA GAT GAA GTC GAT GAA ACA TAC GTC CCC AAA (1610)

501 510 514 516 520 530
glu phe asn ala glu thr phe thr phe his ala asp lle cys thr leu ser glu lys glu arg aln lle lys lys ala leu val
GAG TTT AAT CCT GAA ACA TTC ACC TCC GAT ATA TCC ACA CTT TCT GAG AAC GAC AGA CAA ATC AAC AAA ACT GCA CTT GTC (1700)

0079739

4083

-18-

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-19-

4083

7. Nucleotide sequence coding for the prepeptide of human serum albumin, said nucleotide sequence is as follows:

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-18 p r o
Met lys trp val thr phe ile ser leu leu ohe ser
GCTTTTGTCTCTGTCAACCCACAGGCCCTTGGACACA ATG AAC TGC GTC ATT TCC CTT CTT CTC TTT ACC (30)

-1 -6 p r o -1
ser ala tyr ser arg gly val phe arg arg
TCC CCT TAT TCC AGG CCT GTC TTT CGT CCA

8. Nucleotide sequence coding for pro human serum albumin, said nucleotide sequence is as follows:

35	30	25	20	15	10	5
-6 p r o -1 1						
arg gly val phe arg arg asp ala his lys ser glu val ala his arg phe lys asp leu ala his lys asn phe lys						
ACG GGT GTC TTT CGT CCA GAT GCA CAC AAG AGT GAC GTC GCT CAT CGG TTT AAA GAT TTC GCA GAA AAT TTC AAA (170)						
21 ala leu val lle ala phe ala gln tyr leu gln gln oys pro phe glu asp his val lys leu val asn glu val thr glu phe ala						
GCC TTG GTC ATT GCC TTT CCT CAG TAT CCT CAG CAC TGT CCA TTT GAA GAT CAT GTA AAA TTA GTC AAT GCA GTA ACT CAA TTT GCA (260)						
51 53 60 62 70 75 80						
lys thr oys val ala asp glu ser ala glu asp lys ser leu his thr leu phe gly asp lys leu oys thr val ala thr leu						
AAA ACA TGT GTC CCT CAT GAC TCA GCT GAA AAT TGT GAC AAA TCA CCT CAT ACC CTT TTT CGA GAC AAA TTA TGC ACA GTC ACT CCT (350)						
81 90 91 100 101 110						
arg glu thr tyr gly glu met ala asp oys oys ala lys gln glu pro gly arg asn glu oys phe leu qhn his lys asp aso asn pro						
CGT GAA ACC TAT CCT GAA ATG CCT GAC TCC TGT GCA AAA CAA GAA CCT GGC AGA AAT GAA TGC TTC TGC CAA CAC AAA GAT GAC AAC CCA (440)						
111 120 124 130 140						
asn leu pro arg leu val arg pro glu val asp val met cys thr ala phe his asp asn glu qhn thr phe leu lys tyr leu try						
AAC CTC CCC CGA TTC GTC AGA CAT CCT TAC TTT GAT GTC CCT TTC ACT CCT TTT GCT AAA AGG TAT AAA GCT GCT TTT ACA GAA TGT TGC CAA (530)						
141 150 160 168 169 170						
glu ile ala arg arg his pro tyr phe tyr ala pro glu leu phe ala lys arg tyr lys ala phe thr glu oys cys qhn						
GAA ATT GCC AGA AGA CAT CCT CCT TAC TTT TAT GCC CGG GAA CTC CCT TTC ACT CCT TTT GCT AAA AGG TAT AAA GCT GCT TTT ACA GAA TGT TGC CAA (620)						
171 177 180 190						
ala ala asp lys ala ala oys leu leu pro lys leu asp glu leu arg asp glu gln lys ala ser ser ala lys aln arg leu lys cys						
CCT CCT GAT AAA GCT GCC TGC CTC TGT CCA AAG CTC GAT GAA CCT CGG GAT GAA CGG GAC GCT TCG TCT CCC AAA CAC AGA CTC AAC TGT (710)						
201 210 220 230						
ala ser leu gln lys phe gly glu arg ala phe lys ala trp ala val ala arg leu ser gln arg phe ala glu						
CCC ACT CTC CAA AAA TTT CGA GAA AGA CCT TCC GCA GAA GTC AGC AGC TCC AGC AGA TTT CGA TGT GAC AAA GCT GCA (300)						

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9. Nucleotide sequence coding for the pre pro human serum albumin, said nucleotide sequence is as follows:

35	30	25	20	15	10	5
-1 -6 p r o -1 1	-16 p r o -10					
ser ala tyr ser arg gly val phe arg esp ala his lys ser glu val ala his arg phe lys	Met lys trp val tru phe ile ser leu leu phe leu the ser					
TCC CCT TAT TCC AGC GGT TTT CGT CTC AGC TCG ATT GCT CAC GAC AGT GAG CAC GCA GAT GAA ATT TTC AAA (130)	ATC AAC TCC GTA ACC TTT ATT TCC CTT CAT CGG TTT CTC TTT ACC (130)					
21	30	34	40	50		
ala leu val lle ala phe ala gln tyr leu gln gln cys pro phe glu esp his val lys leu val asn glu val thr glu phe ala						
CCC TTC GTC ATT GCC TTT GCT CAC TAT CTC CAG CAC TGT CCA ATT GAT CAT GCA AAA TTA GTC AAT GAA CTA ACT CAA TTT GCA (1260)						
51 53	60	62	70	75	80	
lys thr cys val ala asp glu ser ala glu asn cys esp lys ser leu his thr leu phe gly asp lys leu oys thr val ala thr leu						
AAA ACA TGT CTT CCT GAT GAG TCA GCT GAA ATT TGT GAC AAA TCA CTT CAT ACC CTT TTT CGA CAC AAA TTA TGC ACA GTC ACT CTT (350)						
81	90 91		100 101			
arg glu thr tyr gly glu met ala asp cys cys ala lys gln glu pro gly arg asn glu oys the leu gln his lys asp asn pro						
CCT GAA ACC TAT GCA ATT CCT GAC CTC TGT GCA AAA CAA CCT CCC AGC ATA ATT GAA CAC AAA TAC GAC AAC CAC AAC CAA (460)						
111	120	124	130	140		
asn leu pro arg leu val arg pro glu val met oys thr ala phe his asp asn glu glu thr phe leu lys tyr leu try						
AAC CTC CCC CCA TTC GTC AGA CCA CCT TGT GAT GTC ATG TCC ACT CCT ATT GCA ATT GAA GAC AGC ATT GAA AAA TAC TTA TAT (330)						
141	150		160	168 169 170		
glu lle ala arg esp his pro tyr phe tyr ala pro glu leu leu phe ohe ala lys arg tyr lys ala ala phe thr glu cys cys qin						
GAA ATT GCG AGA CAT CCT TAC ATT GCT TTT GCT CTC ATT GCA ATT GCA ATT GCT ATT AAA AGC TAT AAA GCA TGT TCC CAA (620)						
171	177	180	190	200		
ala ala esp lys ala cys leu leu pro lys leu esp glu leu arg esp glu qly lys ala ser ser ala lys ala gln aru leu lys cys						
GCT CCT GAT AAA CCT GCG TCC CTC AGC TCC GAT GAA ATT CGG CAT GAA ATT CGG GAT GAA GCG AAC GCT TCC TCT GCA ATT GCA (710)						
201					220	230
ala ser leu gln lys phe gly glu arg ala phe lys ala trp ala val ala arg leu ser gln arg phe pro lys ala glu phe ala glu						
CCC ACT CTC CAA AAA TTT GCA GAA AGA CCT TTC AAA GCA TCC GCA CCT CGC CTC AGC CAG AGA ATT GCA TTT GCA GAA (360)						

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10. A nucleotide sequence according to any of claims 6 to 9, in essentially pure form.
11. A DNA transfer vector comprising a nucleotide sequence as defined in claim 5.
- 5 12. A DNA transfer vector according to claim 11, transferred to and replicated in a micro-organism.
13. A DNA transfer vector according to claim 12, which is a plasmid.
14. A DNA transfer vector according to claim 13,
- 10 wherein the plasmid is pBR322 or YEp6.
15. A process for preparing human serum albumin, which comprises culturing a micro-organism according to claim 5.
16. A DNA transfer vector according to any of
- 15 claims 12 to 14, or a process according to claim 15, wherein the micro-organism is a bacterium or yeast.
17. A vector or process according to claim 16, wherein the bacterium or yeast is E. coli or Saccharomyces cerevisiae.

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Restriction Endonuclease Map of Human Serum Albumin cDNA Clones

